



Improving zinc phytoremediation characteristics in *Salix pedicellata* with a new acclimation approach (2020)

Table 1: New acclimation approach for *S. pedicellata* clones to tolerate DI100 (20 mg L<sup>-1</sup> ZnSO<sub>4</sub>).

Acclimation phases	Time (Days)	ZnSO <sub>4</sub> treatment (mg L <sup>-1</sup> of HS: Hoagland's solution)		
		Induced plants		Control plants
		Low concentrations (mg L <sup>-1</sup> )	Cumulated concentrations (mg L <sup>-1</sup> )	
Tolerance induction phase	0	1.0	1.0	HS
	3	1.1	2.1	HS
	6	1.2	3.3	HS
	9	1.4	4.7	HS
	12	1.6	6.3	HS
	15	1.8	8.1	HS
	18	2.0	10.1	HS
	21	2.1	12.2	HS
	24	2.2	14.4	HS
	27	2.4	16.8	HS
	30	2.6	19.4	HS
	33		20.0	HS
Tolerance maintenance phase	39		20.0	HS
	45		20.0	HS
	51		20.0	HS
	57		20.0	HS

Table 2: Two-way ANOVA test to compare the outcome of ZnSO<sub>4</sub> treatments and biomass accumulation between SP-K12 and SP-K20 *S. pedicellata* clones.

Source	Sum of squares	Degree of freedom	Mean squares	F-value	Probability	Significance
<b>Aerial part</b>						
<b>A:Clone</b>	2.98	1	2.98	8.84	0.0075	**
<b>B:ZnSO<sub>4</sub> treatments</b>	4.13	1	4.13	12.25	0.0023	**
<b>AB</b>	38.10	1	38.10	112.97	0.0000	***
<b>Error</b>	6.75	20	0.34			
<b>Total (Corr.)</b>	51.96	23				
<b>Root part</b>						
<b>A:Clone</b>	3375E – 7	1	3375E – 7	0.01	0.9129	
<b>B:ZnSO<sub>4</sub> treatments</b>	0.43	1	0.43	15.79	0.0007	***
<b>AB</b>	1.51	1	1.51	54.68	0.0000	***
<b>Error</b>	0.55	20	0.028			
<b>Total (Corr.)</b>	2.49	23				

The differences were considered significant at  $p = 0.01-0.05$  (\*); highly significant at  $p = 0.001-0.01$  (\*\*); and very highly significant at  $p \leq 0.001$  (\*\*\*).

Table 3: Two-way ANOVA test to compare the effect of DI100 treatments (20 mg L<sup>-1</sup> ZnSO<sub>4</sub>) on biomass and Zn bioaccumulation in the SP-K20 clone after 6 days of the treatment (66<sup>th</sup> day).

Source	Sum of squares	Degree of freedom	Mean squares	F-value	Probability	Signification
<b>Biomass</b>						
<b>A:DI<sub>100</sub> treatments</b>	78.23	1	78.2287	136.13	0.0000	***
<b>B:Time</b>	29.64	1	29.637	51.57	0.0000	***
<b>AB</b>	17.12	1	17.1197	29.79	0.0000	***
<b>Error</b>	11.49	20	0.574677			
<b>Total (Corr.)</b>	136.48	23				
<b>Zn bioaccumulation</b>						
<b>A: DI<sub>100</sub> treatments</b>	8.27E7	1	8.27E7	477.35	0.0000	***
<b>B:Time</b>	1.54E7	1	1.54E7	88.71	0.0000	***
<b>AB</b>	8.84E6	1	8.84E6	51.02	0.0000	***
<b>Error</b>	3.46E6	20	1.73E5			
<b>Total (Corr.)</b>	1.10E8	23				

The differences were considered significant at  $p = 0.01-0.05$  (\*); highly significant at  $p = 0.001-0.01$  (\*\*); and very highly significant at  $p \leq 0.001$  (\*\*\*).

**Source:** <https://www.tandfonline.com/doi/full/10.1080/15226514.2019.1708862>

# Potential of water lettuce (*Pistia stratiotes* L.) for phytoremediation: physiological responses and kinetics of zinc uptake (2020)

Table 1: Zn contents in the roots and shoots in water lettuce (*Pistia stratiotes* L.) subjected to different treatments and culture times.

Initial Conc. (mg L <sup>-1</sup> )	Metal content in the biomass (mg kg <sup>-1</sup> )				
		24 h	48 h	72 h	168 h
<b>0.7</b>	Root	0.69 ± 0.117	0.73 ± 0.041	0.75 ± 0.136	1.20 ± 0.159
	Shoot	0.17 ± 0.021	0.17 ± 0.011	0.16 ± 0.016	0.40 ± 0.032
<b>1.8</b>	Root	5.96 ± 6.246	6.79 ± 6.795	8.57 ± 8.891	7.66 ± 7.659
	Shoot	0.44 ± 0.022	0.50 ± 0.024	0.65 ± 0.032	1.01 ± 0.015
<b>18.0</b>	Root	23.26 ± 0.708	28.19 ± 1.025	32.75 ± 2.536	46.00 ± 0.979
	Shoot	1.41 ± 0.169	2.40 ± 0.281	2.56 ± 0.092	7.29 ± 0.218
<b>180.0</b>	Root	88.03 ± 0.838	118.21 ± 0.693	105.12 ± 0.397	167.82 ± 0.365
	Shoot	1.41 ± 0.083	5.80 ± 0.021	8.16 ± 0.021	21.58 ± 0.021

Table 2: Percentage Zn removal expressed according to the dry biomass (g) of water lettuce plants subjected to increasing doses of contamination at different culture times.

Initial Conc. (mg.L <sup>-1</sup> )	% Zn removal from the solution				
		24 h	48 h	72 h	168 h
<b>0.7</b>	Root	14.2 ± 1.072	12.0 ± 0.505	16.2 ± 0.315	23.9 ± 0.885
	Shoot	9.6 ± 0.068	9.3 ± 0.265	9.7 ± 0.235	20.4 ± 1.098
	Total	23.8 ± 1.140	21.3 ± 0.504	25.9 ± 0.160	44.3 ± 2.211
<b>1.8</b>	Root	33.1 ± 0.904	40.1 ± 0.981	48.0 ± 2.307	53.7 ± 0.119
	Shoot	4.8 ± 0.022	6.8 ± 0.066	9.2 ± 0.107	18.0 ± 0.702
	Total	37.8 ± 1.853	46.9 ± 0.956	57.2 ± 3.873	71.7 ± 0.767
<b>18.0</b>	Root	14.6 ± 0.378	15.6 ± 0.027	14.9 ± 0.727	28.2 ± 0.474
	Shoot	1.6 ± 0.025	3.5 ± 0.224	4.6 ± 0.087	9.6 ± 0.297
	Total	16.2 ± 0.353	19.1 ± 0.248	19.5 ± 1.234	37.8 ± 0.301
<b>180.0</b>	Root	4.2 ± 0.066	5.4 ± 0.066	3.6 ± 0.113	6.8 ± 0.053
	Shoot	0.6 ± 0.030	0.7 ± 0.045	1.4 ± 0.054	2.4 ± 0.095
	Total	4.8 ± 0.095	6.1 ± 0.164	5.0 ± 0.079	9.2 ± 0.096

Average of three repetitions ± standard deviation.

Table 3. Chlorophyll a (Chl a). chlorophyll b (Chlb). total chlorophyll (C total). and carotenoid (Cx + c) contents. the Cha/Chb ratio. Cttotal/Cx + c ratio and the FCI in water lettuce leaves (*Pistia stratiotes* L.) subjected to increasing doses of Zn at different culture times.

Dependent Variable	Independent Variable	Regression models	Regression coefficient			
$As_{Root}$	$As_{Soil}$ , pH, EC, OM, CEC,	$Y_1 = -1.412 + 0.025 \times As_{Soil}^{**} + 0.493 \times pH^* + 0.018 \times EC + 0.135$	<b>0.96</b>	<b>0.945</b>	<b>64.975</b>	<b>0.0001</b>
	Al%, Fe%	$\times OM^{**} - 0.095 \times CEC - 1.331 \times Al\%^* + 1.63 \times Fe\%^*$				
$As_{Straw}$	$As_{Soil}$ , pH, EC, OM, CEC,	$Y_2 = 0.738 + 0.004 \times As_{Soil}^{**} - 0.134 \times pH + 0.005 \times EC + 0.06 \times$	<b>0.953</b>	<b>0.936</b>	<b>55.59</b>	<b>0.0001</b>
	Al%, Fe%	$OM + 0.013 \times CEC - 0.38 \times Al\%^* + 0.352 \times Fe\%^*$				
$As_{Grain}$	$As_{Soil}$ , pH, EC, OM, CEC	$Y_3 = 0.554 + 0.003 \times As_{Soil}^{**} - 0.127 \times pH + 0.004 \times EC + 0.055 \times$	<b>0.94</b>	<b>0.918</b>	<b>42.415</b>	<b>0.0001</b>
	Al%, Fe%	$OM + 0.004 \times CEC - 0.246 \times Al\%^* + 0.195 \times Fe\%^*$				

Average of three repetitions  $\pm$  standard deviation.

\*Chl a/Chl b = ratio between chlorophyll a and chlorophyll b contents.

\*Ctotal/Cx + c = ratio between total chlorophyll (chlorophyll a + b) and carotenoid contents.

\*FCI: Falker chlorophyll index.

Table 4: Kinetic parameters of Zn uptake by water lettuce (*Pistia stratiotes* L.) cultivated in nutrient solutions with different concentrations of the element.

Solution Conc. (mg L <sup>-1</sup> )	Kinetic parameters	
	$K_m$ (μmol L <sup>-1</sup> )	$V_{max}$ (μmol g <sup>-1</sup> h <sup>-1</sup> )
<b>1.8</b>	1.590 $\pm$ 0.035	0.080 $\pm$ 0.018
<b>18.0</b>	61.240 $\pm$ 2.065	0.189 $\pm$ 0.008

Average of three repetitions  $\pm$  standard deviation.

Table 5. Variation in biomass and root/shoot ratio of water lettuce (*Pistia stratiotes* L.) after 93 h of culture in nutrient solution contaminated with different doses of Zn.

Solution concentration (mg L <sup>-1</sup> )	Biomass (grams)		
	Initial	Final	Root/shoot ratio
<b>1.8</b>	42.03 $\pm$ 0.494	34.86 $\pm$ 0.985	0.43 $\pm$ 0.049
<b>18</b>	45.98 $\pm$ 0.630	41.91 $\pm$ 0.890	0.63 $\pm$ 0.042

Average of three repetitions  $\pm$  standard deviation.

Source: <https://www.tandfonline.com/doi/full/10.1080/15226514.2020.1725868>

## Phytotoxicity Increase Induced by Zinc Accumulation in *Cichorium intybus* (2020)

Table 1: Mean leaf areas (square inch) of *Cichorium* after growing in natural and Zn-contaminated soils for 90 days

Days	Natural soil	Zn-contaminated soil	
	Non-contaminated	1200 mg kg <sup>-1</sup>	2400 mg kg <sup>-1</sup>
15	2.56 <sup>a</sup> ± 0.05	2.64 <sup>a</sup> ± 0.02	2.61 <sup>a</sup> ± 0.02
30	4.10 <sup>b</sup> ± 0.09	3.99 <sup>b</sup> ± 0.01	2.67 <sup>b</sup> ± 0.04
45	8.35 <sup>c</sup> ± 0.01	4.21 <sup>c</sup> ± 0.05	2.31 <sup>c</sup> ± 0.02
60	9.56 <sup>d</sup> ± 0.10	3.54 <sup>de</sup> ± 0.04	2.28 <sup>cd</sup> ± 0.03
75	14.63 <sup>ef</sup> ± 0.12	3.53 <sup>cd</sup> ± 0.02	1.20 <sup>e</sup> ± 0.01
90	14.75 <sup>f</sup> ± 0.04	3.32 <sup>f</sup> ± 0.05	0.95 <sup>f</sup> ± 0.01

Means with different letters in the vertical columns are significantly different by t-test at a 95% confidence level

Table 2: Mean leaf biomasses (g) of *Cichorium* after growing in natural and Zn-contaminated soils for 90 days

Days	Natural soil	Zn-contaminated soil	
	Non-contaminated	1200 mg kg <sup>-1</sup>	2400 mg kg <sup>-1</sup>
15	0.23 <sup>a</sup> ± 0.01	0.24 <sup>a</sup> ± 0.04	0.23 <sup>a</sup> ± 0.01
30	0.41 <sup>b</sup> ± 0.03	0.40 <sup>b</sup> ± 0.02	0.41 <sup>b</sup> ± 0.02
45	0.83 <sup>c</sup> ± 0.01	0.42 <sup>bc</sup> ± 0.04	0.37 <sup>c</sup> ± 0.02
60	1.62 <sup>d</sup> ± 0.04	0.66 <sup>d</sup> ± 0.03	0.36 <sup>cd</sup> ± 0.04
75	1.78 <sup>e</sup> ± 0.01	0.58 <sup>e</sup> ± 0.03	0.25 <sup>ae</sup> ± 0.01
90	1.96 <sup>f</sup> ± 0.05	0.49 <sup>f</sup> ± 0.03	0.19 <sup>f</sup> ± 0.02

Means with different letters in the vertical columns are significantly different by t-test at a 95% confidence level

Table 3: Mean leaf length (LL in inch) and mean root length (RL in square inch) of *Cichorium* after growing in natural and Zn-contaminated soils for 90 days

Days	Natural soil		Zn-contaminated soil			
	Non-contaminated		1200 mg kg <sup>-1</sup>		2400 mg kg <sup>-1</sup>	
	LL	RL	LL	RL	LL	RL
15	4.53 <sup>a</sup> ± 0.04	3.94 <sup>a</sup> ± 0.05	4.30 <sup>a</sup> ± 0.08	2.44 <sup>a</sup> ± 0.05	4.09 <sup>a</sup> ± 0.12	1.97 <sup>a</sup> ± 0.05
30	5.30 <sup>b</sup> ± 0.02	5.67 <sup>b</sup> ± 0.10	4.47 <sup>b</sup> ± 0.05	3.35 <sup>b</sup> ± 0.04	3.65 <sup>b</sup> ± 0.04	1.98 <sup>ab</sup> ± 0.02
45	5.32 <sup>bc</sup> ± 0.03	13.86 <sup>c</sup> ± 1.02	5.07 <sup>c</sup> ± 0.02	5.59 <sup>c</sup> ± 0.07	3.15 <sup>c</sup> ± 0.20	2.05 <sup>c</sup> ± 0.05
60	6.46 <sup>d</sup> ± 0.07	17.17 <sup>d</sup> ± 0.08	4.53 <sup>d</sup> ± 0.02	6.30 <sup>d</sup> ± 0.12	3.11 <sup>cd</sup> ± 0.05	1.99 <sup>ad</sup> ± 0.02
75	6.90 <sup>e</sup> ± 0.03	19.40 <sup>e</sup> ± 0.03	4.16 <sup>e</sup> ± 0.02	3.58 <sup>e</sup> ± 0.08	3.05 <sup>de</sup> ± 0.01	1.88 <sup>e</sup> ± 0.01
90	7.05 <sup>f</sup> ± 0.00	20.47 <sup>f</sup> ± 0.05	4.15 <sup>ef</sup> ± 0.04	2.76 <sup>f</sup> ± 0.01	2.85 <sup>f</sup> ± 0.03	1.87 <sup>ef</sup> ± 0.02

Means with different letters in the vertical columns are significantly different by t-test at a 95% confidence level

Table 4: Concentrations of Zn ( $\text{mg kg}^{-1}$ ) in Cichorium leaves after growing in natural and Zn-contaminated soils for 90 days

Days	Natural Soil	Zn-contaminated soil	
	Non-contaminated	1200 $\text{mg kg}^{-1}$	2400 $\text{mg kg}^{-1}$
15	2.31 <sup>a</sup> $\pm$ 0.75	334.0 <sup>a</sup> $\pm$ 25.6	584.5 <sup>a</sup> $\pm$ 43.4
30	5.35 <sup>b</sup> $\pm$ 1.05	401.9 <sup>ab</sup> $\pm$ 1.05	620.6 <sup>a</sup> $\pm$ 57.4
45	9.43 <sup>c</sup> $\pm$ 2.44	481.6 <sup>bc</sup> $\pm$ 35.0	1136 <sup>b</sup> $\pm$ 68.2
60	15.6 <sup>d</sup> $\pm$ 1.50	529.5 <sup>c</sup> $\pm$ 30.1	1995 <sup>c</sup> $\pm$ 37.6
75	25.0 <sup>e</sup> $\pm$ 0.54	700.3 <sup>d</sup> $\pm$ 55.6	2028 <sup>c</sup> $\pm$ 32.9
90	37.5 <sup>f</sup> $\pm$ 3.89	809.7 <sup>d</sup> $\pm$ 4.75	2232 <sup>d</sup> $\pm$ 16.7

Means with different letters in the vertical columns are significantly different by t-test at a 95% confidence level

**Source:** <https://link.springer.com/article/10.1007/s00128-020-02960-4>

# Zinc tolerant plant growth promoting bacteria alleviates phytotoxic effects of zinc on maize through zinc immobilization (2020)

Table 1: Soil sample sites and chemical properties of experimented soil.

Place	Satellite location	EC <sup>a</sup> (dS/m)	pH <sup>a</sup>	OC (g/kg)	Av. N (kg/ha)	Av. P (kg/ha)	Av. K (kg/ha)	DTPA-Zn (mg/kg)
Mochia, Zawar	24° 21' 37.6" N 73° 41' 45.3" E	0.57	7.19	0.55	94.82	20.20	199.36	35.99
Balaria, Zawar	24° 35' 38.8" N 73° 75' 21.1" E	0.62	7.25	0.60	81.09	18.22	169.44	39.99

<sup>a</sup>1:2 soil to water ratio, OC, organic carbon; Av. N, available nitrogen (Kjeldahl digestion); Av. P, available phosphorus (Olsen's P<sub>2</sub>O<sub>5</sub>); Av. K, available potassium (ammonium acetate extractable K<sub>2</sub>O).

Table 2: Biochemical characterization of zinc tolerant bacteria.

Strain name	Starch hydrolysis	Citrate utilization	Nitrate reduction	Gelatin liquefaction	Catalase activity	Oxidase
ZTB 15	–	+	–	+	+	–
ZTB 24	+	+	+	–	+	–
ZTB 28	+	+	–	–	+	+
ZTB 29	+	+	–	–	+	–

+, Positive; –, negative.

Table 3: Effect of Zn concentration on biosorption of Zn by ZTB.

Strain name	Concentration of Zn (mg/L) in the supernatant after biosorption by ZTB after 72 h		% Biosorption of Zn by ZTB after 72 h	
	Media with 20 mg/L Zn	Media with 40 mg/L Zn	Media with 20 mg/L Zn	Media with 40 mg/L Zn
ZTB 15	1.508 ± 0.196 <sup>a</sup>	2.598 ± 0.252 <sup>a</sup>	92.46	93.51
ZTB 24	3.285 ± 0.020 <sup>c</sup>	9.586 ± 0.121 <sup>c</sup>	83.58	76.04
ZTB 28	1.831 ± 0.050 <sup>b</sup>	3.851 ± 0.059 <sup>b</sup>	90.85	91.87
ZTB 29	1.825 ± 0.309 <sup>ab</sup>	3.597 ± 0.252 <sup>b</sup>	90.88	91.01

Data is presented as means of 3 replicates ± SD (standard deviation). The Mean value followed by same letter in column of each treatment is not significant difference at  $p = 0.05$  by Tukey–Kramer HSD test.

Table 4: Plant growth promoting activities of ZTB.

PGPR activity	ZTB strains			
	ZTB15	ZTB24	ZTB28	ZTB29
IAA production (µg/mL)	4.83 ± 0.02	4.32 ± 0.040	8.03 ± 0.02	12.54 ± 0.07
ACC deaminase activity	++	+	++	+
Ammonia production (µg/mL)	1.42 ± 0.23	1.49 ± 0.56	1.48 ± 0.18	1.45 ± 0.86
HCN production	–	–	–	–
GA <sub>3</sub> (µg/mL)	28.20 ± 1.31	60.60 ± 1.50	40.86 ± 1.23	28.10 ± 1.01
Phosphate solubilization index	4.60 ± 0.10	3.45 ± 0.10	4.10 ± 0.20	3.85 ± 0.04
Potassium solubilization index	4.20 ± 0.05	6.30 ± 0.05	6.33 ± 0.03	8.00 ± 0.10
Silica solubilization index	2.23 ± 0.02	2.90 ± 0.01	3.52 ± 0.01	2.30 ± 0.01
Phytase production index	12.12 ± 0.01	11.42 ± 0.01	7.50 ± 0.02	11.42 ± 0.01
Siderophore index (Z/C)	2.08 ± 0.01	1.66 ± 0.01	1.11 ± 0.01	2.00 ± 0.60

+, Positive; ++, medium positive; +++, high positive; –, negative; Data is presented as means of 3 replicates ± SD (standard deviation).

**Table 5: In vitro studies on the effect of zinc tolerant bacteria on growth and biomass of maize seedling under Zn stress conditions (1,000 mg Zn/kg planting mixture).**

Treatment details	Average shoot length (cm)	Average root length (cm)	Average root number	Average leaf number	Total chlorophyll (µg/mL)
<b>T1: control without Zn and ZTB inoculation</b>	11.50 ± 0.93 <sup>c</sup>	38.50 ± 4.03 <sup>b</sup>	10.52 ± 0.98 <sup>cd</sup>	6.00 ± 1.0 <sup>a</sup>	34.14 ± 4.14 <sup>b</sup>
<b>T2: control with Zn and without ZTB inoculation</b>	8.90 ± 1.03 <sup>bc</sup>	36.50 ± 3.20 <sup>b</sup>	10.13 ± 0.86 <sup>d</sup>	5.00 ± 0.58 <sup>a</sup>	32.83 ± 4.91 <sup>b</sup>
<b>T3: with Zn and ZTB15 inoculation</b>	13.2 ± 1.47 <sup>b</sup>	47.23 ± 2.07 <sup>a</sup>	13.33 ± 1.32 <sup>bc</sup>	5.30 ± 1.15 <sup>a</sup>	47.10 ± 4.0 <sup>a</sup>
<b>T4: with Zn and ZTB24 inoculation</b>	13.26 ± 1.25 <sup>b</sup>	48.56 ± 2.22 <sup>a</sup>	14.33 ± 1.25 <sup>b</sup>	6.30 ± 0.58 <sup>a</sup>	47.10 ± 3.77 <sup>a</sup>
<b>T5: with Zn and ZTB28 inoculation</b>	16.59 ± 0.90 <sup>a</sup>	52.96 ± 3.04 <sup>a</sup>	17.67 ± 1.23 <sup>a</sup>	6.67 ± 1.15 <sup>a</sup>	57.87 ± 3.99 <sup>a</sup>
<b>T6: with Zn and ZTB29 inoculation</b>	13.85 ± 1.10 <sup>ab</sup>	50.23 ± 1.94 <sup>a</sup>	14.33 ± 1.08 <sup>b</sup>	5.30 ± 1.15 <sup>a</sup>	48.67 ± 4.26 <sup>a</sup>
<b>CD at 5%</b>	2.21	4.42	2.21	2.01	6.73
<b>CV%</b>	14.08	7.97	13.61	28.93	12.44

Data are recorded after 30 days of germination; data is presented as means of 4 replicates ± SD (standard deviation). The Mean value followed by same letter in column of each treatment is not significant difference at  $p = 0.05$  by Tukey–Kramer HSD test.

**Table 6: In vitro studies on the effect of ZTB on stress related enzymes of maize seedling under Zn stress conditions (1,000 mg Zn /kg planting mixture).**

Treatment details	SOD (unit/mg) fresh weight	POD (µmole/min/g)	PAL (µmole/min/g)	Catalase (µmole/min/g)	PPO (µmole/min/g)
<b>T1: control without Zn and ZTB inoculation</b>	0.21 ± 0.02 <sup>f</sup>	1.80 ± 0.18 <sup>g</sup>	0.0203 ± 0.002 <sup>fg</sup>	18.50 ± 0.41 <sup>d</sup>	0.0127 ± 0.001 <sup>d</sup>
<b>T2: control with Zn and without ZTB inoculation</b>	0.27 ± 0.02 <sup>ab</sup>	1.95 ± 0.30 <sup>ab</sup>	0.0213 ± 0.001 <sup>ab</sup>	19.23 ± 0.25 <sup>a</sup>	0.0141 ± 0.001 <sup>c</sup>
<b>T3: with Zn and ZTB15 inoculation</b>	0.36 ± 0.03 <sup>cde</sup>	2.82 ± 0.20 <sup>bc</sup>	0.0233 ± 0.006 <sup>g</sup>	20.92 ± 1.95 <sup>cd</sup>	0.0170 ± 0.002 <sup>a</sup>
<b>T4: with Zn and ZTB24 inoculation</b>	0.39 ± 0.03 <sup>bc</sup>	2.27 ± 0.25 <sup>ef</sup>	0.0283 ± 0.002 <sup>cdef</sup>	22.58 ± 1.26 <sup>c</sup>	0.0163 ± 0.002 <sup>b</sup>
<b>T5: with Zn and ZTB28 inoculation</b>	0.33 ± 0.03 <sup>def</sup>	2.20 ± 0.25 <sup>ef</sup>	0.0314 ± 0.001 <sup>bc</sup>	21.17 ± 2.05 <sup>cd</sup>	0.0174 ± 0.001 <sup>a</sup>
<b>T6: with Zn and ZTB29 inoculation</b>	0.37 ± 0.03 <sup>cd</sup>	2.71 ± 0.25 <sup>cd</sup>	0.0301 ± 0.005 <sup>cd</sup>	26.53 ± 1.51 <sup>ab</sup>	0.0176 ± 0.002 <sup>a</sup>
<b>CD at 5%</b>	0.050	0.460	0.010	1.430	0.001
<b>CV%</b>	8.61	11.15	14.10	3.65	4.42

\*Value is mean of 4 replicates. The Mean value followed by same letter in column of each treatment is not significant difference at  $p = 0.05$  by Tukey–Kramer HSD test.

**Table 7: In vitro studies on the effect of ZTB on Zn accumulation in maize seedling under Zn stress conditions (1,000 mg Zn /kg planting mixture).**

Treatment details	Zn concentration in shoot (µg/g fresh weight)	Zn concentration in root (µg/g fresh weight)
<b>T1: control without Zn and ZTB inoculation</b>	65.01 ± 5.0 <sup>d</sup>	46.03 ± 6.5 <sup>e</sup>
<b>T2: control with Zn and without ZTB inoculation</b>	632.64 ± 6.0 <sup>a</sup>	487.90 ± 11.5 <sup>a</sup>
<b>T3: with Zn and ZTB15 inoculation</b>	356.28 ± 5.1 <sup>b</sup>	299.70 ± 10.1 <sup>b</sup>
<b>T4: with Zn and ZTB24 inoculation</b>	335.31 ± 7.6 <sup>bc</sup>	280.20 ± 5.5 <sup>bc</sup>
<b>T5: with Zn and ZTB28 inoculation</b>	333.12 ± 7.5 <sup>c</sup>	262.20 ± 7.0 <sup>c</sup>
<b>T6: with Zn and ZTB29 inoculation</b>	339.57 ± 7.1 <sup>bc</sup>	218.70 ± 4.45 <sup>d</sup>
<b>CD at 5%</b>	2.44	6.22
<b>CV%</b>	0.39	1.29

Each value is mean of 4 replicates. The Mean value followed by same letter in column of each treatment is not significant difference at  $p = 0.05$  by Tukey–Kramer HSD test.

**Source:** <https://www.nature.com/articles/s41598-020-70846-w>



## Exposure of biosynthesized nanoscale ZnO to *Brassica Juncea* crop plant: morphological, biochemical and molecular aspects (2020)

**Table 1: Number of interactions in the binding site determined through docking between  $\alpha$ -amylase and Zinc acetate with their respective type of interaction.**

Index	Entry	Gold score. Fitness	Interacting residues	Ionic/Hydrogen bond interactions number
1	Structure2D_CID_11192   dock1	36.2783	Tyr155, Gln158, Lys209, Tyr238	6
2	Structure2D_CID_11192   dock3	36.0536	Tyr155, Gln158, Lys209, Tyr238	6
3	Structure2D_CID_11192   dock2	35.4436	Tyr155, Gln158, Lys209, Tyr238	6

**Table 2: Effect of ZnO NPs on different crop plants.**

Plants	NP Concentration in soil/water	NP size(nm)	Effects
<i>Macrotyloma uniflorum</i>	2–100 mg/L	50	Delayed germination time
<i>Fagopyrum esculentum</i>	10–2,000 mg/L	<50 nm	Decreased the biomass content
<b>Bean</b>	500 mg/kg	<100 nm	Reduced root growth
<b>Soybean</b>	500 mg/kg	<100 nm	Ceased seed production
<i>Glycine max</i>	2,000 and 4,000 mg/L	55–70	Genotoxic
<b>Lettuce</b>	10 mg/kg	41–48	Enhanced the photosynthesis and biomass
<i>Cyamopsis tetragonoloba</i>	10 mg/kg	67	Increased its biomass, shoot-root length, length, chlorophyll content, and total soluble leaf protein
<i>Triticum aestivum</i>	20 mg/L	<100 nm	Increased grain yield and increase in shoot dry weight.
<i>Arachis hypogaea</i> L.	400 and 1000 mg/L	25–100	Improvement in the germination rate and seedling vigor index
<b>Tomato and egg plants</b>	1.0 mg/mL	38–46	Boost plant defence and yield
<i>Brassica nigra</i>	500 to 1500 mg/L	<100 nm	Reduced seed germination and seedling growth
<i>Brassica napus</i>	10 to 250 mg/L	155 ± 10	Chlorosis at high concentration
<i>Brassica juncea</i>	10–30 µg/ml	11 nm	Increased germination and chlorophyll biosynthesis rate along with low ROS production at 20 µg/ml.
			At 30 µg/ml germination rate, chlorophyll biosynthesis decreases and ROS production increases.

**Source:** <https://www.nature.com/articles/s41598-020-65271-y>

# Impact of selenium, zinc and their interaction on key enzymes, grain yield, selenium, zinc concentrations, and seedling vigor of biofortified rice (2020)

**Table 1 Chemical properties of experimental dry soil**

No	Index	Content	Method
2	pH	5.8	Potentiometry (NYT 1377–2007)
3	Organic carbon (g/kg of soil)	38.4	Potassium dichromate volumetric method (NYT 1121–6-2006)
4	Extractable nitrogen (mg/kg of soil)	141	Universal extraction colometric method (NYT 1849–2010)
5	Extractable phosphorus (mg/kg of soil)	28.1	Universal extraction colometric method (NYT 1849–2010)
6	Extractable potassium (mg/kg of soil)	113	Universal extraction colometric method (NYT 1849–2010)
7	Total selenium (mg/kg of soil)	0.24	Fluorescence spectrophotometry (NYT1104–2006)
8	Bioavailable selenium (mg/kg of soil)	0.026	Extraction with 0.016 M potassium dihydrogen phosphate
9	Total zinc (mg/kg of soil)	2.75	(NY/T 890–2004)
10	Available zinc (mg/kg of soil)	1.48	Extraction with DTPA (diethylenetriaminepentaacetic acid)
11	Sulfur (mg/kg of soil)	2.75	Extraction with phosphate-acetic acid solution (NY/T 1121-14-2006)
12	Cadmium (mg/kg of soil)	0.13	Graphite furnace atomic absorption spectrophotometry (GB/T17141–1997)

**Table 2: Treatments and Rates of zinc (Zn) and selenium (Se) nutrients soil application**

No	Treatments	Zn	Se
		(mg/kg of soil)	(mg/kg of soil)
T1	Control	0	0
T2	Zn <sub>5</sub>	5	0
T3	Zn <sub>10</sub>	10	0
T4	Zn <sub>15</sub>	15	0
T5	Se(1 mg/kg)	0	1
T6	Se(1 mg/kg) + Zn <sub>5</sub>	5	1
T7	Se(1 mg/kg) + Zn <sub>10</sub>	10	1
T8	Se(1 mg/kg) + Zn <sub>15</sub>	15	1

**Table 3: Total chlorophyll, carotenoids, superoxide dismutase (SOD), and catalase (CAT) activity of R725 rice genotype as affected by single selenium, zinc, and combined selenium-zinc (Se-Zn) addition**

No	Treatments	Total chlorophyll content (mg/g of fresh weight)	Carotenoids (mg/g of fresh weight)	SOD (U/g of fresh weight)	CAT (nmol/g /min/g of fresh weight)
T1	Control	2.85 a	1.81 d	574.99 a	520.2 ab
T2	Zn <sub>5</sub>	2.85 a	2.07 bc	691.11 a	555.39 a
T3	Zn <sub>10</sub>	3.16 a	2.11 bc	696.00 a	590.58 a
T4	Zn <sub>15</sub>	3.27 a	2.71 a	641.01 a	465.12 ab
T5	Se(1 mg/kg)	3.26 a	1.83 d	299.21 b	250.92 d
T6	Se(1 mg/kg) + Zn <sub>5</sub>	3.25 a	2.35 bc	308.31 b	304.47 cd
T7	Se(1 mg/kg) + Zn <sub>10</sub>	3.5 a	2.23 bc	318.24 b	406.98 bc
T8	Se(1 mg/kg) + Zn <sub>15</sub>	3.28 a	2.49 ab	272.9 b	550.80 a
	SEM±	0.26	0.15	35.37	36.67

Data in the column with the same letter are not significantly different at the 0.05 level by least significant difference test; SEM ± stand for means of standard error

**Table 4: Grain yield and total dry matter of R725 rice genotype as affected by single selenium (Se), zinc (Zn), and combined selenium-zinc (Se-Zn) addition**

No	Treatments	Grain yield (g/pot)	Total dry matter (g/pot)
<b>T1</b>	Control	108.73 cd	221.07 b
<b>T2</b>	Zn <sub>5</sub>	125.27 ab	259.67 a
<b>T3</b>	Zn <sub>10</sub>	131.6 ab	264.84 a
<b>T4</b>	Zn <sub>15</sub>	126.67 ab	257.63 a
<b>T5</b>	Se(1 mg/kg)	100.33 d	211.77 b
<b>T6</b>	Se(1 mg/kg) + Zn <sub>5</sub>	119.93 bc	245.40 a
<b>T7</b>	Se(1 mg/kg) + Zn <sub>10</sub>	124.6 ab	258.47 a
<b>T8</b>	Se(1 mg/kg) + Zn <sub>15</sub>	133.57a	259.73 a
	SEm±	3.58	5.44

Data in the column with the same letter are not significantly different at the 0.05 level by least significant difference test; SEm ± stand for means of standard error

**Table 5: Zinc concentration in different parts of grain of R725 rice genotype as affected by single selenium, zinc (Zn), and combined selenium-zinc (Se-Zn) addition**

No	Treatments	Zinc concentration (mg/kg) in			
		Grain	Husk	Brown rice	Polished rice
<b>T1</b>	Control	79.82 b	43.08 bc	36.74 a	15.22 a
<b>T2</b>	Zn <sub>5</sub>	70.98 b	38.28 bc	32.70 a	18.29 a
<b>T3</b>	Zn <sub>10</sub>	77.68 b	35.97 c	41.71 a	14.61 a
<b>T4</b>	Zn <sub>15</sub>	82.04 b	41.63 bc	40.38 a	18.99 a
<b>T5</b>	Se(1 mg/kg)	101.46 a	62.18 a	39.28 a	15.81 a
<b>T6</b>	Se(1 mg/kg) + Zn <sub>5</sub>	84.07 ab	50.79 ab	33.28 a	15.50 a
<b>T7</b>	Se(1 mg/kg) + Zn <sub>10</sub>	79.62 b	38.29 bc	41.33 a	17.53 a
<b>T8</b>	Se(1 mg/kg) + Zn <sub>15</sub>	70.32 b	38.23 bc	32.09 a	17.06 a
	SEm±	5.22	3.47	2.99	1.54

Data in the column with the same letter are not significantly different at the 0.05 level by least significant difference test; SEm ± stand for means of standard error

**Table 6: Selenium concentration in different parts of grain of R725 rice genotype as affected by single selenium (Se), zinc (Zn), and combined selenium-zinc (Se-Zn) addition**

No	Treatments	Selenium concentration (mg/kg) in			
		Grain	Husk	Brown rice	Polished rice
<b>T1</b>	Control	0.09 c	0.051 b	0.039 d	0.028 d
<b>T2</b>	Zn <sub>5</sub>	0.085 c	0.047 b	0.037 d	0.034 d
<b>T3</b>	Zn <sub>10</sub>	0.094 c	0.053 b	0.041 d	0.036 d
<b>T4</b>	Zn <sub>15</sub>	0.085 c	0.051 b	0.034 d	0.033 d
<b>T5</b>	Se(1 mg/kg)	0.59 b	0.195 a	0.400 c	0.337 c
<b>T6</b>	Se(1 mg/kg) + Zn <sub>5</sub>	0.775a	0.288 a	0.487 ab	0.408 bc
<b>T7</b>	Se(1 mg/kg) + Zn <sub>10</sub>	0.8493a	0.283 a	0.567 a	0.533 a
<b>T8</b>	Se(1 mg/kg) + Zn <sub>15</sub>	0.716 ab	0.222 a	0.494 ab	0.448 ab
	SEm±	0.036	0.026	0.024	0.017

Data in the column with the same letter are not significantly different at the 0.05 level by least significant difference test; SEm ± stand for means of standard error

**Table 7: Germination%, length of coleoptile, and shoot and root as well as plant dry weight grown from seeds of R725 rice genotype harvested from single selenium, zinc (Zn), and combined selenium-zinc (Se-Zn) treatments**

No	Treatments	Germination%	Length (mm) of			Plant Dry Weight (g/30 plants)
			<b>Coleoptile</b>	<b>Shoot</b>	<b>Root</b>	
<b>T1</b>	Control	96.67ab	1.42 ab	6.84 d	8.90b	0.72 a
<b>T2</b>	Zn <sub>5</sub>	96.67 ab	1.45 ab	7.47 cd	9.06 b	0.75 a
<b>T3</b>	Zn <sub>10</sub>	94.00 b	1.36 ab	7.48 cd	8.84 b	0.72 a
<b>T4</b>	Zn <sub>15</sub>	96.00 ab	1.62 a	8.24 bc	9.18 b	0.71 a
<b>T5</b>	Se(1 mg/kg)	94.00 b	1.32 b	7.25 d	8.41 b	0.66 a
<b>T6</b>	Se(1 mg/kg) + Zn <sub>5</sub>	96.67 ab	1.22 b	7.30d	9.11 b	0.64 a
<b>T7</b>	Se(1 mg/kg) + Zn <sub>10</sub>	98.67 a	1.62 a	8.50 b	10.84 a	0.71 a
<b>T8</b>	Se(1 mg/kg) + Zn <sub>15</sub>	98.00 ab	1.36 ab	9.86 a	12.04 a	0.71 a
	SEm±	1.86	0.12	0.37	0.64	0.05

Data in the column with the same letter are not significantly different at the 0.05 level by least significant difference test; SEm ± stand for means of standard error

**Source:** <https://link.springer.com/article/10.1007%2Fs11356-020-08202-8>

## Effects of exogenous zinc on the photosynthesis and carbonic anhydrase activity of millet (*Setaria italica* L.) (2020)

Table 1: Effect of exogenous zinc on the pigment content of millet leaves. Values are means  $\pm$  SE (n = 3). Different letters in the same column indicate significant difference at the  $p < 0.05$  level by Duncan's new multiple range test. Different concentrations of Zn solution (0, 20, 40, 60, 80, and 100 mg L<sup>-1</sup>) were sprayed at the seedling stage of millet and recorded as CK, Zn<sub>1</sub>, Zn<sub>2</sub>, Zn<sub>3</sub>, Zn<sub>4</sub>, and Zn<sub>5</sub>, respectively.

Cultivar	Treatment	Chl <i>a</i> [mg g <sup>-1</sup> (FM)]	Chl <i>b</i> [mg g <sup>-1</sup> (FM)]	Chl ( <i>a+b</i> ) [mg g <sup>-1</sup> (FM)]	Carotenoid [mg g <sup>-1</sup> (FM)]
<b>Zhangzagu 10</b>	CK	9.66 $\pm$ 0.18c	3.77 $\pm$ 0.17b	13.43 $\pm$ 0.11c	1.40 $\pm$ 0.04ab
<b>Zn1</b>	10.13 $\pm$ 0.05d	4.07 $\pm$ 0.12cd	14.21 $\pm$ 0.17e	1.47 $\pm$ 0.04c	
<b>Zn2</b>	10.40 $\pm$ 0.09e	4.21 $\pm$ 0.03d	14.62 $\pm$ 0.10f	1.47 $\pm$ 0.03c	
<b>Zn3</b>	9.99 $\pm$ 0.07d	3.97 $\pm$ 0.11c	13.96 $\pm$ 0.04d	1.42 $\pm$ 0.01bc	
<b>Zn4</b>	9.35 $\pm$ 0.10b	3.47 $\pm$ 0.09a	12.82 $\pm$ 0.19b	1.36 $\pm$ 0.05ab	
<b>Zn5</b>	9.14 $\pm$ 0.10a	3.27 $\pm$ 0.11a	12.41 $\pm$ 0.10a	1.35 $\pm$ 0.02a	
<b>Jingu 21</b>	CK	7.36 $\pm$ 0.11c	2.38 $\pm$ 0.16bc	9.74 $\pm$ 0.16c	1.31 $\pm$ 0.09ab
<b>Zn1</b>	8.02 $\pm$ 0.12d	2.60 $\pm$ 0.32c	10.62 $\pm$ 0.25d	1.38 $\pm$ 0.13ab	
<b>Zn2</b>	8.36 $\pm$ 0.09e	2.95 $\pm$ 0.13d	11.32 $\pm$ 0.12e	1.42 $\pm$ 0.03b	
<b>Zn3</b>	7.86 $\pm$ 0.05d	2.56 $\pm$ 0.14c	10.41 $\pm$ 0.19d	1.34 $\pm$ 0.05ab	
<b>Zn4</b>	7.02 $\pm$ 0.09b	2.17 $\pm$ 0.07ab	9.19 $\pm$ 0.16b	1.27 $\pm$ 0.01a	
<b>Zn5</b>	6.66 $\pm$ 0.09a	1.95 $\pm$ 0.17a	8.61 $\pm$ 0.09a	1.26 $\pm$ 0.08a	

Table 2: Effect of exogenous zinc on photosynthetic gas-exchange parameters of millet. Values are means  $\pm$  SE (n = 3). Different letters in the same column indicate significant difference at the  $p < 0.05$  level by Duncan's new multiple range test. Different concentrations of Zn solution (0, 20, 40, 60, 80, and 100 mg L<sup>-1</sup>) were sprayed at the seedling stage of millet and recorded as CK, Zn<sub>1</sub>, Zn<sub>2</sub>, Zn<sub>3</sub>, Zn<sub>4</sub>, and Zn<sub>5</sub>, respectively. PN – net photosynthetic rate; gs – stomatal conductance; E – transpiration rate; Ci – intercellular CO<sub>2</sub> concentration.

Cultivar	Treatment	PN [ $\mu$ mol m <sup>-2</sup> s <sup>-1</sup> ]	gs [mmol m <sup>-2</sup> s <sup>-1</sup> ]	E [mmol m <sup>-2</sup> s <sup>-1</sup> ]	Ci [ $\mu$ mol mol <sup>-1</sup> ]
<b>Zhangzagu 10</b>	CK	6.95 $\pm$ 0.13c	54.92 $\pm$ 0.56c	2.388 $\pm$ 0.051c	212.05 $\pm$ 1.05d
<b>Zn1</b>	7.50 $\pm$ 0.21d	56.90 $\pm$ 0.34e	2.489 $\pm$ 0.019e	195.18 $\pm$ 1.28b	
<b>Zn2</b>	8.07 $\pm$ 0.15e	58.24 $\pm$ 0.27f	2.555 $\pm$ 0.020f	188.22 $\pm$ 2.54a	
<b>Zn3</b>	7.38 $\pm$ 0.11d	56.42 $\pm$ 0.44d	2.463 $\pm$ 0.023d	203.63 $\pm$ 4.39c	
<b>Zn4</b>	6.51 $\pm$ 0.13b	53.71 $\pm$ 0.29b	2.314 $\pm$ 0.016b	217.23 $\pm$ 3.26e	
<b>Zn5</b>	6.06 $\pm$ 0.22a	52.35 $\pm$ 0.27a	2.243 $\pm$ 0.014a	223.44 $\pm$ 2.49f	
<b>Jingu 21</b>	CK	5.76 $\pm$ 0.13c	50.56 $\pm$ 0.84c	2.154 $\pm$ 0.071c	249.58 $\pm$ 1.33d
<b>Zn1</b>	6.42 $\pm$ 0.12e	53.23 $\pm$ 0.39e	2.284 $\pm$ 0.029d	221.47 $\pm$ 0.82b	
<b>Zn2</b>	7.04 $\pm$ 0.13f	55.02 $\pm$ 0.20f	2.365 $\pm$ 0.011e	207.34 $\pm$ 1.73a	
<b>Zn3</b>	6.23 $\pm$ 0.14d	52.61 $\pm$ 0.34d	2.265 $\pm$ 0.013d	234.20 $\pm$ 4.38c	
<b>Zn4</b>	5.11 $\pm$ 0.15b	48.05 $\pm$ 0.12b	2.037 $\pm$ 0.021b	260.70 $\pm$ 3.12e	
<b>Zn5</b>	4.83 $\pm$ 0.18a	46.24 $\pm$ 0.33a	1.937 $\pm$ 0.036a	267.47 $\pm$ 2.31	

### Source:

[https://www.researchgate.net/publication/340753868\\_Effects\\_of\\_exogenous\\_zinc\\_on\\_the\\_photosynthesis\\_and\\_carbonic\\_anhydrase\\_activity\\_of\\_millet\\_Setaria\\_italica\\_L](https://www.researchgate.net/publication/340753868_Effects_of_exogenous_zinc_on_the_photosynthesis_and_carbonic_anhydrase_activity_of_millet_Setaria_italica_L)

Phytotoxicity of nano-zinc oxide to tomato plant (*Solanum lycopersicum* L): Zn uptake, stress enzymes response and influence on non-enzymatic antioxidants in fruits (2019)

Table 1: Effect of n-ZnO on chlorophyll contents (mg/g fw) of *Solanum lycopersicum* L. (30 day exposure)

	Chl-a (30 day exposure)	Chl-b (30 day exposure)	T-Chl
<b>Control</b>	597±157a	533±138a	1130±221a
<b>300 mg n-ZnO/kg</b>	387±82b	163±41b	550±223b
<b>600 mg n-ZnO/kg</b>	217±65c	103±15b	320±56c
<b>1000 mg n-ZnO/kg</b>	300±20c	190±10b	490±26c

(90 day exposure)

	Chl-a (30 day exposure)	Chl-b (30 day exposure)	T-Chl
<b>Control</b>	607±85 <sup>a</sup>	150±51b	750±87ab
<b>300 mg n-ZnO/kg</b>	657±50a	190±43a	847±91a
<b>600 mg n-ZnO/kg</b>	433±35b	227±40a	660±75ab
<b>1000 mg n-ZnO/kg</b>	263±69c	110±10c	367±72c

**Note:** Values are means ± SD. Mean with the same letter(s) along the same column are not statistically different at p<0.05 by Turkey.

The nano-zinc oxide significantly affected the chlorophyll contents at early stage of the growth. Chl-a, -b and T-Chl at 30 days were all significantly reduced compared to control for all n-ZnO-treatments. The treatments caused reduction of Chl-a, b and T-Chl by at least 54.3%, 99.6% and 105.4%, respectively at 30 day exposure. The 90-day exposure effect of n-ZnO treatment on chlorophyll contents showed that the treatments did alter the contents of Chl-a, and T-Chl at ≤ 600 mg n-ZnO/kg.

**Source:** <https://www.sciencedirect.com/science/article/abs/pii/S2352186418304681>

## Comparison study of zinc nanoparticles and zinc sulphate on wheat growth: from toxicity and zinc biofortification (2019)

**Table 1: Effects of Zn treatments on Zn concentration in different parts of wheat**

Treatments	Grain (mg kg <sup>-1</sup> )		Glume (mg kg <sup>-1</sup> )		Stem (mg kg <sup>-1</sup> )		Leaf (mg kg <sup>-1</sup> )		Root (mg kg <sup>-1</sup> )	
(mg kg <sup>-1</sup> )	ZnO	ZnSO <sub>4</sub>	ZnO	ZnSO <sub>4</sub>	ZnO	ZnSO <sub>4</sub>	ZnO	ZnSO <sub>4</sub>	ZnO	ZnSO <sub>4</sub>
	NPs		NPs		NPs		NPs		NPs	
<b>Control</b>	18.3e	18.3cd	10.4d	10.4d	8.7d	10.4e	6.6d	8.7e	15.5e	<b>15.5d</b>
<b>10</b>	22.6de	20.6c	15.2c	12.7d	12.3cd	12.7e	8.2cd	12.3d	18.4d	<b>16.2d</b>
<b>20</b>	27.1d	25.9bc	16.9c	15.6c	15.9c	15.6d	9.6c	15.9c	20.2cd	<b>18.7cd</b>
<b>50</b>	43.6c	29.6b	12.7cd	18.5c	17.5c	18.5c	13.5b	17.5c	23.4c	<b>20.3c</b>
<b>100</b>	50.4b	31.1b	17.0c	25.3b	21.1b	25.3b	14.0b	21.1b	35.2b	<b>22.6c</b>
<b>200</b>	52.4b	35.4b	28.0b	25.0b	22.3b	25.0b	11.9bc	22.3b	39.0b	<b>28.8b</b>
<b>1000</b>	<b>60.4a</b>	<b>44.2a</b>	<b>37.7a</b>	<b>31.0a</b>	<b>39.7a</b>	<b>31.0a</b>	<b>20.1a</b>	<b>39.7a</b>	<b>82.7a</b>	<b>39.8a</b>

**Note:** Totally different lower case letters followed with values in the same column indicate significant differences between treatments ( $p < 0.05$ ).

Zn can be accumulated in all tissues through soil as shown by results from the pot trial. All plant organs showed increased Zn content with the increase in treatment concentrations. The concentration of Zn in grains increased by 3.3 times and 2.4 times for ZnO NPs and ZnSO<sub>4</sub> at 1000 mg kg<sup>-1</sup>. On the contrary, ZnSO<sub>4</sub> was more effective at increasing leaf Zn than ZnO NPs, which increased remarkably from 41% to 356% and 24% to 205%, showed an average rate of 147% and 95% for ZnSO<sub>4</sub> and ZnO NPs, respectively. Du et al. (2011) reported the similar results that Zn accumulations were significantly enhanced in different tissues treated with ZnO NPs.

**Source:** <https://www.sciencedirect.com/science/article/pii/S0045653519306149>

## Comparison study of zinc nanoparticles and zinc sulphate on wheat growth: From toxicity and zinc biofortification (2019)

Table 1: Effects of Zn treatments on grain yield, aboveground biomass and harvest index of wheat

Treatments (mg kg <sup>-1</sup> )	Grain Yield (g pot <sup>-1</sup> )		Above ground Biomass (g pot <sup>-1</sup> )		Harvest Index (%)	
	ZnO NPs	ZnSO <sub>4</sub>	ZnO NPs	ZnSO <sub>4</sub>	ZnO NPs	ZnSO <sub>4</sub>
<b>Control</b>	12.5b	12.5c	33.3cd	33.3d	37.5ab	<b>37.5a</b>
<b>10</b>	13.2ab	14.5b	37.1c	42.4b	35.6ab	<b>34.2ab</b>
<b>20</b>	18.6a	19.4a	54.4a	57.4a	34.2ab	<b>33.8ab</b>
<b>50</b>	19.5a	18.6a	48.8ab	57.1a	39.9a	<b>32.6b</b>
<b>100</b>	16.8b	18.5a	44.8b	52.1a	37.5ab	<b>35.5ab</b>
<b>200</b>	15.4b	13.6bc	47.2ab	37.0c	32.6b	<b>36.8ab</b>
<b>1000</b>	<b>10.4c</b>	<b>8.5d</b>	<b>29.3d</b>	<b>23.9e</b>	<b>35.5ab</b>	<b>35.6ab</b>

In terms of the harvest index means, at 50 mg kg<sup>-1</sup>, the harvest index increased by 6% for ZnO NPs, while all treatments with ZnSO<sub>4</sub> reduced harvest index.

**Source:** <https://www.sciencedirect.com/science/article/pii/S0045653519306149>

## Effects of zinc fertilizer amendments on yield and grain zinc concentration under controlled environment conditions. (2018)

Table 1: Summary of Zn fertilizer treatments in pot experiment.

Treatment	Zn Application Method	Zn Application Rate (kg Zn ha <sup>-1</sup> )
<b>Control</b>	N/A <sup>a</sup>	0.000
<b>ZnSO<sub>4</sub></b>	Soil	2.500
<b>7% Zn lignosulphonate</b>	Foliar	0.246
<b>9% Zn chelated with EDTA</b>	Foliar	0.246
<b>9% Zn chelated with EDTA</b>	Soil	0.246

Table 2: Effects of various forms of Zn fertilize on grain and straw yield (g pot<sup>-1</sup>) of three lentil cultivars.

Fertilizer	Cultivar	Yield (g pot <sup>-1</sup> ) <sup>a</sup>	
		Grain	Straw
Control	CDC Maxim	1.47 a	1.97 c
	CDC Invincible	1.43 a	1.92 c
	CDC Impower	1.29 a	3.00 a
Soil ZnSO <sub>4</sub>	CDC Maxim	1.45 a	1.92 c
	CDC Invincible	1.38 a	1.79 c
	CDC Impower	1.37 a	2.93 a
7% Zn foliar lignosulphonate	CDC Maxim	1.32 a	2.19 bc
	CDC Invincible	1.35 a	1.91 c
	CDC Impower	1.43 a	2.71 ab
9% Zn foliar EDTA chelated	CDC Maxim	1.36 a	1.84 c
	CDC Invincible	1.31 a	1.86 c
	CDC Impower	1.35 a	2.78 a
9% Zn soil EDTA chelated	CDC Maxim	1.52 a	1.85 c
	CDC Invincible	1.35 a	1.98 c
	CDC Impower	1.33 a	2.72 ab
SEM <sup>b</sup>		0.08	0.12
Statistical Analysis		P values	
Fertilizer effect		0.828	0.579
Cultivar effect		0.309	<0.0001
Fertilizer × cultivar interaction effect		0.662	0.334

<sup>a</sup>Means with the same letter in the same column are not significantly different ( $P > .05$ ) as determined by multi-treatment comparisons using the Tukey-Kramer method.

<sup>b</sup>SEM=standard error of mean.



Table 3: Effects of various forms of Zn fertilizer on grain and straw Zn concentration (mg Zn kg<sup>-1</sup>) of three lentil cultivars.

Fertilizer	Cultivar	Zn Concentration (mg Zn kg <sup>-1</sup> ) <sup>a</sup>	
		Grain	Straw
Control	CDC Maxim	36.7 a	29.5 a
	CDC Invincible	38.2 a	31.4 a
	CDC Impower	33.3 a	31.5 a
Soil ZnSO <sub>4</sub>	CDC Maxim	36.2 a	24.4 a
	CDC Invincible	35.3 a	29.1 a
	CDC Impower	33.7 a	32.2 a
7% Zn foliar lignosulphonate	CDC Maxim	41.0 a	30.1 a
	CDC Invincible	38.4 a	30.3 a
	CDC Impower	34.9 a	31.5 a
9% Zn foliar EDTA chelated	CDC Maxim	41.6 a	33.2 a
	CDC Invincible	32.8 a	31.9 a
	CDC Impower	36.9 a	31.6 a
9% Zn soil EDTA chelated	CDC Maxim	37.3 a	32.8 a
	CDC Invincible	39.1 a	30.6 a
	CDC Impower	43.5 a	30.6 a
SEM <sup>b</sup>		4.53	2.21
Statistical Analysis		P values	
Fertilizer effect		0.708	0.353
Cultivar effect		0.719	0.569
Fertilizer × cultivar interaction effect		0.859	0.536

<sup>a</sup>Means with the same letter in the same column are not significantly different ( $P > 0.05$ ) as determined by multi-treatment comparisons using the Tukey-Kramer method.

<sup>b</sup>SEM = standard error of mean.

Table 4: Zinc removal (mg Zn pot<sup>-1</sup>) in lentil cultivars amended with different forms of Zn fertilizer.

Cultivar	Zn Uptake and Removal (μg Zn pot <sup>-1</sup> ) <sup>a</sup>		
	Straw	Grain	Total
CDC Maxim	58.7 b	54.2 a	112.9 b
CDC Invincible	58.1 b	50.1 a	108.2 b
CDC Impower	89.9 a	49.6 a	139.4 a
SEM <sup>b</sup>	2.92	3.00	4.54
P value	<0.0001	0.49	<.0001

<sup>a</sup>Means with the same letter in the same column are not significantly different ( $P > 0.05$ ) as determined by multi-treatment comparisons using the Tukey-Kramer method.

<sup>b</sup>SEM = standard error of mean.

Source: <https://www.tandfonline.com/doi/full/10.1080/01904167.2018.1462386>

# Zinc effect on growth rate, chlorophyll, protein and mineral contents of hydroponically grown mungbeans plant (*Vigna radiata*) (2017)

**Table 1: Plant height (cm) on dry weight basis in mungbean varieties at different concentrations of Zn in solution culture.**

Zn treatments	V1	V2	V3	V4	Mean ± St. dv
Control	<b>19.60b</b>	<b>19.93d</b>	<b>19.53bc</b>	<b>13.03e</b>	<b>18.02b 3.33</b>
1 $\mu$ M	<b>22.94a</b>	<b>22.60a</b>	<b>22.70a</b>	<b>20.73 cd</b>	<b>22.24a 1.02</b>
2 $\mu$ M	<b>23.18a</b>	<b>23.00a</b>	<b>23.20a</b>	<b>21.03bc</b>	<b>22.60a 1.05</b>
Mean ± St.d v	<b>21.91a</b>	<b>21.84a</b>	<b>21.81a</b>	<b>20.27b</b>	
	<b>2.00</b>	<b>1.67</b>	<b>1.99</b>	<b>4.53</b>	

V1 = Ramazan, V2 = Swat mungI, V3 = NM92, V4 = KMI.  
St. dv = standard deviation.

The mean followed by similar letter (s) are not significantly different at P= 0.05

**Table 2: Chlorophyll contents (mgkg<sup>-1</sup>) on dry weight basis in mungbean varieties at different concentrations of Zn in solution culture.**

Zn treatments	V1	V2	V3	V4	Mean ± St. dv
Control	<b>35.7f</b>	<b>73.45de</b>	<b>93.12 cd</b>	<b>105.93c</b>	<b>78.55b 30.63</b>
1 $\mu$ M	<b>36.81f</b>	<b>145.30b</b>	<b>210.82a</b>	<b>221.01a</b>	<b>153.5a 84.71</b>
2 $\mu$ M	<b>64.54e</b>	<b>146.07b</b>	<b>210.57a</b>	<b>226.08a</b>	<b>153.5a 84.71</b>
Mean ± St.d v	<b>45.69c</b>	<b>123.6b</b>	<b>171.5a</b>	<b>184.4a</b>	
	<b>16.34</b>	<b>41.71</b>	<b>67.88</b>	<b>67.95</b>	

V1 = Ramazan, V2 = Swat mungI, V3 = NM92, V4 = KMI.  
St. d = standard deviation.

The mean followed by similar letter (s) are not significantly different at P = 0.05.

**Table 3: Percent crude protein (dry weight basis) in mungbean varieties at different concentrations of Zn in solution culture.**

Zn treatments	V1	V2	V3	V4	Mean ± St. dv
Control	<b>12.90f</b>	<b>11.76f</b>	<b>13.95ef</b>	<b>11.54f</b>	<b>12.54c 1.11</b>
1 $\mu$ M	<b>13.12f</b>	<b>16.45de</b>	<b>17.62bcd</b>	<b>18.12 cd</b>	<b>16.08b 2.25</b>
2 $\mu$ M	<b>20.54ab</b>	<b>22.86a</b>	<b>20.99a</b>	<b>22.05abc</b>	<b>21.61a 1.05</b>
Mean ± St.d v	<b>15.52a</b>	<b>17.02a</b>	<b>18.02a</b>	<b>17.24a</b>	
	<b>4.35</b>	<b>5.57</b>	<b>3.52</b>	<b>4.32</b>	

V1 = Ramazan, V2 = Swat mungI, V3 = NM92, V4 = KMI.  
St. d = standard deviation.

The mean followed by similar letter (s) are not significantly different at P = 0.05.

**Source :** <https://www.sciencedirect.com/science/article/pii/S1878535213002050>

## Silicon addition to soybean (*Glycine max L.*) plants alleviate zinc deficiency (2016)

Table 1: Zinc content ( $\mu\text{mol plant}^{-1}$ ) at the three sampling times (M1, M2 and M3) after Zn removal from the NS. Values within a column followed by different letters differ significantly ( $P < 0.05$ , Duncan test).

Treatment	Leaves Zn ( $\mu\text{mol leaves}^{-1}$ )			Stems Zn ( $\mu\text{mol stems}^{-1}$ )			Root Zn ( $\mu\text{mol root}^{-1}$ )		
	M1	M2	M3	M1	M2	M3	M1	M2	M3
Zn 0 Si (0.0–0.0)	0.44 d	0.68 c	0.84 d	0.057 c	0.165 b	0.270 c	0.12 d	0.24 b	0.37 b
Zn 10 Si (0.0–0.0)	1.29 ab	1.25 a	5.26 a	0.075 bc	0.052 c	0.414 a	0.65 a	0.22 bc	2.73 a
Zn 0 Si (0.5–0.5)	1.12 bc	0.92 b	1.35 b	0.083 bc	0.129 b	0.358 ab	0.14 d	0.15 d	0.49 b
Zn 0 Si (1.0–1.0)	1.05 c	1.05 b	1.19 bc	0.096 ab	0.250 a	0.415 a	0.27 bc	0.37 a	0.38 b
Zn 0 Si (0.5–0.0)	1.36 a	1.36 a	1.07 c	0.075 bc	0.165 b	0.278 c	0.18 cd	0.16 cd	0.56 b
Zn 0 Si (1.0–0.0)	1.03 c	0.96 b	1.21 bc	0.118 a	0.176 b	0.334 bc	0.38 b	0.17 cd	0.43 b

Source: <https://www.sciencedirect.com/science/article/pii/S0981942816302753>

## Zinc Fertilization Under Optimum Soil Moisture Condition Improved the Aromatic Rice Productivity (2016)

Table 1: Effect of zinc fertilization and irrigation regimes on maximum LAI

Treatments	Zn1	Zn2	Zn3	Zn4	Zn5
Shekhupura (Site 1)					
I <sub>1</sub>	5.29	5.55	5.72	5.89	5.88
I <sub>2</sub>	5.74	5.92	6.16	6.1	6.11
I <sub>3</sub>	5.76	5.83	6.01	6.48	6.82
I <sub>4</sub>	5.82	6.06	6.98	7.24	7.3
I <sub>5</sub>	6.4	6.83	6.96	7.46	7.51
Sargodha (Site 2)					
I <sub>1</sub>	3.38	3.5	3.78	3.87	3.64
I <sub>2</sub>	3.57	3.69	3.87	3.92	3.96
I <sub>3</sub>	3.96	4.28	4.58	4.86	5.14
I <sub>4</sub>	3.85	4.28	4.58	5.13	5.59
I <sub>5</sub>	4.2B	4.6	5.2	5.28	5.08

LSD (Site 1) = 0.2, LSO (Site 2) = 0.13, Zn1 (0 kg ha<sup>-1</sup>), Zn2 (8 kg ha<sup>-1</sup>), Zn3 (10 kg ha<sup>-1</sup>), Zn4 (12 kg ha<sup>-1</sup>), Zn5 (14 kg ha<sup>-1</sup>), I<sub>1</sub> (6 irrigations), I<sub>2</sub> (8 irrigations), I<sub>3</sub> (10 irrigations), I<sub>4</sub> (12 irrigations), I<sub>5</sub> (14 irrigations)

Table 2: Effect of zinc fertilization and irrigation regimes on LAD

Treatments	Zn <sub>0</sub>	Zn <sub>1</sub>	Zn <sub>2</sub>	Zn <sub>3</sub>	Zn <sub>4</sub>
<b>Shekhupura (Site 1)</b>					
I <sub>1</sub>	2.96.45	307.5	316.39	325.54	330.94
I <sub>2</sub>	301.2	320.82	328.6	336.26	347.54
I <sub>3</sub>	313.47	330.17	339	373.88	400.55
I <sub>4</sub>	323	346	376.1	407.86	424.83
I <sub>5</sub>	345.42	378.89	394.81	422.38	411.31
<b>Sargodha (Site 2)</b>					
I <sub>1</sub>	192.6	209.57	223.97	234.06	232.49
I <sub>2</sub>	210.5	219.58	236.81	244.38	251.15
I <sub>3</sub>	233.22	260.19	276.37	293.58	309.7
I <sub>4</sub>	235.43	259.37	278.72	313.69	337.61
I <sub>5</sub>	25.93	278.89	307.86	320.72	312.97

LSD (Site 1) = 0.2. LSD (Site 2) = 0.13, Zn<sub>0</sub> (0 kg ha<sup>-1</sup>), Zn<sub>1</sub> (8 kg ha<sup>-1</sup>), Zn<sub>2</sub> (10 kg ha<sup>-1</sup>), Zn<sub>3</sub> (12 kg ha<sup>-1</sup>), Zn<sub>4</sub> (14 kg ha<sup>-1</sup>), I<sub>1</sub> (6 irrigations), I<sub>2</sub> (8 irrigations), I<sub>3</sub> (10 irrigations), I<sub>4</sub> (12 irrigations), I<sub>5</sub> (14 irrigations)

Table 3: Effect of zinc fertilization and irrigation regimes on NAR (g m<sup>-2</sup> day<sup>-1</sup>)

Treatments	Zn <sub>0</sub>	Zn <sub>1</sub>	Zn <sub>2</sub>	Zn <sub>3</sub>	Zn <sub>4</sub>
<b>Shekhupura (Site 1)</b>					
I <sub>1</sub>	3.34	3.4	3.49	3.6	3.77
I <sub>2</sub>	3.79	3.74	3.67	3.58	3.4
I <sub>3</sub>	3.86	3.64	4.06	3.67	3.66
I <sub>4</sub>	4.6	4.53	4.37	4.15	4.18
I <sub>5</sub>	4.11	3.78	3.75	3.62	3.68
<b>Sargodha (Site 2)</b>					
I <sub>1</sub>	2.92	2.98	3.07	3.18	3.35
I <sub>2</sub>	3.37	3.32	3.25	3.16	2.98
I <sub>3</sub>	3.44	3.22	3.64	3.25	3.24
I <sub>4</sub>	4.18	4.11	3.95	3.73	3.76
I <sub>5</sub>	3.14	3.36	3.33	3.2	3.26

LSD (Site 1) = 0.2, LSD (Site 2) = 0.13, Zn<sub>0</sub> (0 kg ha<sup>-1</sup>), Zn<sub>1</sub> (8 kg ha<sup>-1</sup>), Zn<sub>2</sub> (10 kg ha<sup>-1</sup>), Zn<sub>3</sub> (12 kg ha<sup>-1</sup>), Zn<sub>4</sub> (14 kg ha<sup>-1</sup>), I<sub>1</sub> (6 irrigations), I<sub>2</sub> (8 irrigations), I<sub>3</sub> (10 irrigations), I<sub>4</sub> (12 irrigations), I<sub>5</sub> (14 irrigations)

Table 4: Effect of zinc fertilization and irrigation regimes on total tillers per hill

Treatments	Zn <sub>0</sub>	Zn <sub>1</sub>	Zn <sub>2</sub>	Zn <sub>3</sub>	Zn <sub>4</sub>
<b>Shekhupura (Site 1)</b>					
I <sub>1</sub>	14.27	14.54	15.64	16.94	18.29
I <sub>2</sub>	16.34	17.56	17.58	17.57	17.18
I <sub>3</sub>	17.42	17.66	18.43	18.37	18.8
I <sub>4</sub>	19.03	19.42	19.62	19.81	20.19
I <sub>5</sub>	18.95	18.5	18.77	19.1	18.92
<b>Sargodha (Site 2)</b>					
I <sub>1</sub>	13.55	13.76	14.19	14.59	14.85
I <sub>2</sub>	14.69	15.2	15.58	15.86	16.41
I <sub>3</sub>	15.79	16.32	16.53	16.88	17.21
I <sub>4</sub>	15.81	16.46	17.43	17.87	18.04
I <sub>5</sub>	16.53	17.63	17.76	17.92	17.91

LSD (Site 1) = 0.2, LSD (Site 2) = 0.13, Zn<sub>0</sub> (0 kg ha<sup>-1</sup>), Zn<sub>1</sub> (8 kg ha<sup>-1</sup>), Zn<sub>2</sub> (10 kg ha<sup>-1</sup>), Zn<sub>3</sub> (12 kg ha<sup>-1</sup>), Zn<sub>4</sub> (14 kg ha<sup>-1</sup>), I<sub>1</sub> (6 irrigations), I<sub>2</sub> (8 irrigations), I<sub>3</sub> (10 irrigations), I<sub>4</sub> (12 irrigations), I<sub>5</sub> (14 irrigations)

Source: <https://www.cabdirect.org/cabdirect/abstract/20163306772>